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Handheld Enzymatic Luminescent Biosensor for Rapid Detection of Heavy Metals in Water Samples

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Abstract: Enzymatic luminescent systems are a promising tool for rapid detection of heavy metals ions for water quality assessment. Nevertheless, their widespread use is limited by the lack of test procedure automation and available sensitive handheld luminometers. Herein we describe integration of disposable microfluidic chips for bioluminescent enzyme-inhibition based assay with a handheld luminometer, which detection system is based on a thermally stabilized silicon photomultiplier (SiPM). Microfluidic chips were made of poly(methyl methacrylate) by micro-milling method and sealed using a solvent bonding technique. The composition of the bioluminescent system in microfluidic chip was optimized to achieve higher luminescence intensity and storage time. Results indicate that developed device provided comparable sensitivity with bench-scale PMT-based commercial luminometers. Limit of detection for copper (II) sulfate reached 2.5 mg/l for developed biosensor. Hereby we proved the concept of handheld enzymatic optical biosensors with disposable chips for bioassay. The proposed biosensor can be used as an early warning field-deployable system for rapid detection of heavy metals salts and other toxic chemicals, which affect bioluminescent signal of enzymatic reaction.

Keywords: Chemical Measurements; Silicon Photomultiplier; Optical Biosensor; Bioassay; Microfluidics; Luciferase; Bioluminescence

1. Introduction

Significant problem affecting human health is the global pollution of water sources by various types of organic and inorganic toxic substances [1]. Analytical methods and tools that allow rapid and inexpensive monitoring of the environment are required. Traditional analytical methods such as mass spectrometry and chromatography are highly expensive and therefore their application on an ongoing basis or in the field is limited. In this context, biosensors appear as suitable alternatives or complementary analytical tools [2].

A biosensor can be defined as an independently integrated receptor transducer device, which is capable of providing selective quantitative or semi-quantitative analytical information using a biological recognition element [3]. Molecular biosensors use enzymes, immunosystems, tissues, organelles or whole cells as recognition elements to detect chemical compounds usually by electrical, thermal or optical signals [4]. Biosensors are more favorable, reliable, accurate, cost effective, and easy to use compared to other conventional lab-based detection techniques due to their portability, reusability, real-time response, high specificity and selectivity [5]. It is worth noting that the combination of all these qualities in one device is still one of the challenges in the development

of field deployable biosensors [6]. Despite these difficulties, for more than 40 years of their history, biosensors have found wide commercial application in a variety of fields, such as medical tests, food quality analyses, biothreat, environmental protection, etc. [7].

When using biological objects as recognition elements, it is always worth considering the fact that in nature there are no two absolutely identical organisms or cells. Such biodiversity introduces a certain error in the measurement. The use of individual molecules, such as proteins, avoids this problem and comes as close as possible to the accuracy and unambiguity of chemical measurements [8]. A necessary condition for this is the presence of such a detector that does not affect the measuring element of the biosensor when measured.

Among various types of biosensors, the optical ones tend to be the most promising, especially in the field of environmental pollution control. This is due to their high sensitivity, no need for extensive sample preparation, multitarget sensing, the possibility of compact design, and in most cases label-free detection, except for quantum dots and fluorescence-based methods [9,10]. Bioluminescence-based biorecognition elements have great potential for producing ccost-effective and compact optical biosensors because of their low cost [11,12]. A bacterial coupled enzyme system NAD(P)H:FMN-oxidoreductase and luciferase (Red+Luc) can be used as such kind of biorecognition element for development of express assays of water quality [13,14]. Luciferase catalyzes the oxidation of long-chain aliphatic aldehydes involving reduced flavin mononucleotide; one of the products of this reaction is a quantum of light in the blue-green spectrum. To provide luciferase with reduced flavin mononucleotide, the luciferase reaction is coupled with the reaction catalyzed by NAD(P)H:FMN-oxidoreductase [15–17].

The interaction of toxicants with the coupled enzyme system Red+Luc leads to the change of such bioluminescence parameters as the luminescence intensity, delay in the output of the reaction to the maximum luminescence emission and alteration of chemical reaction constants. This approach does not provide information on the chemical composition of the harmful compounds in the sample. However, it makes it possible to assess the effect of these unknown compounds on the biological system and provides a warning signal that chemical analysis is required to identify the substance.

One of the trends in the development of optical biosensors in the field of environmental pollution control and early warning systems is integration of microelectronics and microfluidics into optical biosensors for miniaturization of optical biorecognition elements [18]. This leads to the development of disposable chips [19,20], which are more suitable for use with portable biosensors and protect samples from cross-contamination. Recently we have introduced disposable luciferase-based microfluidic chips to perform enzymatic assay [21–23]. These chips can be used for rapid assay of water pollution. However, there was a lack of portable luminometers suitable for work with microfluidic chips.

The development of portable luminometers imposes restrictions on their size, including the size of the optical detection system. Silicon photomultipliers (SiPM) seems to be promissing photodetectors for portable biosensors, because of the several advantages [24,25]: low operating voltage (smaller than 50 V), insensitivity to magnetic fields, compact dimensions, and immunity to damage from light overexposure, high gain (10^6). They are chiper than compact photomultiplier tubes and relatively new to the market. SiPM potential is being currently studied for biomedical application [26], compact imaging systems [27,28], etc. by a lot of research groups in the world [29,30]. Sensitivity of the SiPM detector depends on the ambient temperature, which can be overcome by using a photodetector cooling system [31].

Here we introduce the integration of luciferase-based microfluidic chips with a portable luminometer in order to develop handheld enzymatic luminescent biosensor for rapid detection of heavy metals in water samples. To develop a biosensor, it was necessary (i) to design a portable luminometer, (ii) to optimize the composition of bioluminescent system in microfluidic chip.

2. Materials and Methods

2.1. Reagents

The following reagents were used: FMN (CHEBI: 17621, Serva, Germany), reduced nicotinamide adenine dinucleotide (NADH) (CHEBI: 16908, Gerbu, Germany), ethanol (CHEBI:16236, Merk, Germany), tetradecanal (CHEBI:84067, Merck, Germany), starch from potato (CHEBI:28017, Sigma-Aldrich, USA), gelatin from porcine skin (CHEBI:5291, Sigma-Aldrich, USA), potassium phosphate buffer with pH 7.0 (CHEBI:63036, Fluka, Sweden), PMMA (CHEBI:61369, SoftPlast, Russia), 1,2-dichloroethane (Soyuzhimprom, Russia) and acetone (Vekton, Russia). Lyophilized preparations of purified enzymes were produced at the Laboratory of Nanobiotechnology and Bioluminescence of the Institute of Biophysics SB RAS (Krasnoyarsk, Russia). One vial of preparation contained 0.5 mg of luciferase EC 1.14.14.3 (*Photobacterium leiognathi*) from recombinant strain of *Escherichia coli* and 0.18 activity units of NAD(P)H:FMN-oxidoreductase EC 1.5.1.29 (*Vibrio fischeri*).

2.2. Biosensor fabrication

The housing of the portable luminometer was designed using a CAD software KOMPAS-3D (Ascon, Russia). Then the housing was manufactured by a micro-milling method with the CNC milling machine Modela MDX-40A (Roland, Japan) and model plastic Necuron 1300 (NECUMER GmbH, Germany). The sample compartment was manufactured by pressing heated polyvinyl chloride (PVC) into the mould, which was made by the same technique as the housing of the device. Disposable microfluidic chips were manufactured in accordance with the technique described in detail earlier [21, 22]. In brief, the body of the chip consisted of two PMMA (polymethylmethacrylate) substrates. One of the substrates contained a channelized surface made by micro-milling [32]. The enzymes and substrates of bioluminescent system Red+Luc were immobilized in the reaction chamber of the chip in the form of dried droplets of starch gel. The chips were sealed at room temperature using solvent bonding technique.

Vinylpolysiloxane A-silicone (Zhermack, Italy) was used as a material for sampler fabrication. A mould for casting was made by micro-milling.

2.3. Signal detection

The SiPM consisted of a PM6650-EB (KETEK, Germany) containing an array of 14272 avalanche photodiodes with individual cell dimensions of $50 \times 50\,\mu$m providing an active area of $6 \times 6\,mm^2$. The signal from the SiPM was amplified by operational amplifier AD8062 (Analog Devices, USA), and then counted by a microcontroller as peaks counts per second expressed as relative luminescence units (RLU). This information was shown on the biosensor display and the data were also transmitted to a personal computer (PC) through a USB interface. The program for micro controller was developed with Oberon programming language [33] using Astrobe (CFB Software, Australia) and O7 (Alexander Shiryaev, Russia) compilers. The program for PC was developed with open-source IDE BlackBox Component Builder (Oberon microsystems AG, Switzerland).

The intensity of bioluminescent signal from microfluidic chip was measured by the developed device and GloMax 20/20 luminometer (Promega, USA) in the mode for kinetics measurement.

2.4. Testing procedure

The testing procedure was as follows. At the first stage the chip was filled with liquid sample using a sampler adapter for microfluidic chips. At the 30th second after the sample introduction mixing with sampler adapter started. Ten pushes on the sampler membrane resulted in uniform distribution of the bioluminescent system reagents in the reaction chamber of the chip. Then the chip was placed in the luminometer, where the luminescence intensity detection was performed. At the final stage the waste chip was removed from the luminometer. Then the procedure was repeated with a new chip.

The activity of the immobilized coupled enzyme system Red+Luc was measured in the following way. At the beginning, we registered the control luminescence intensity of the enzyme system (I_c). For I_c registration, the chip was filled with distilled water sample. The chip was placed in the luminometer. Then the luminescence intensity of the analyzed sample was measured (I_{exp}). For I_{exp} registration, the chip was filled with analyzed liquid sample.

Statistical analysis was performed using t-distribution at a 95% range.

3. Results and discussion

3.1. Design of portable luminometer

The biosensor was composed of three main parts: a compact and portable luminometer, a disposable microfluidic chip and a sampler adapter (Figure 1). The microfluidic chip had the dimensions 14.4 × 40.5 mm, which was less than a traditional glass microscope slide, and contained components of the coupled enzyme system Red+Luc co-immobilized into starch gel [23].

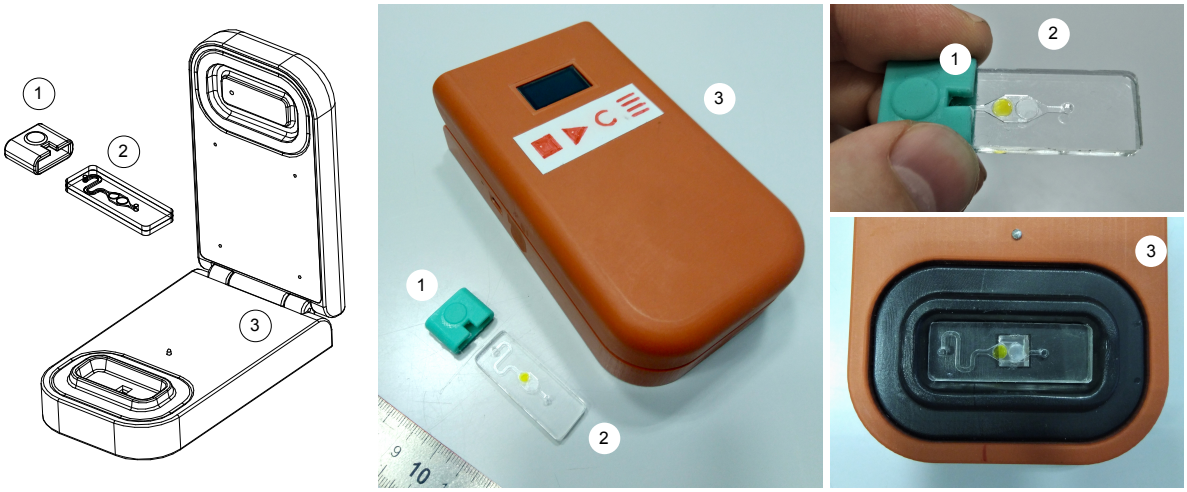

Figure 1. The biosensor consists of a SiPM-based handheld luminometer (3), a disposable microfluidic chips (2) and a sampler adapter for chips (1). The portable luminometer has the capability of autonomous operation, for this purpose it is equipped with a battery, a display and control buttons. The disposable microfluidic chips contain enzymes and substrates of the coupled enzyme bioluminescent system NAD(P)H:FMN-oxidoreductase-luciferase co-immobilized into starch gel

The compact luminometer measured 80 × 140 × 41 cm and weighed less than 300 g. It consisted of a sample compartment, a silicon photomultiplier with a thermal stabilization system, a battery and a microcontroller electronics. All the elements were combined in a hard plastic housing that shielded the internal components from light and dust. The principal design was inspired by the work of Wojciechowski [34]. Because of the nature of the readout method, shielding from ambient light was extremely important. The sample compartment was made from an opaque PVC. The size of the sample compartment measured 41 × 15 × 5 mm, thus making it possible to use not only special microfluidic chips but also other planar samples (test strips, slices, etc.).

The designed software proved to be versatile and user-friendly, plotting in real-time during measurement and choosing the signal integration time. The information about the current measurement was duplicated on the biosensor display. All the measurements could be started and stopped using the buttons built into the housing of the biosensor.

The portable luminometer had the possibility of autonomous operation due to the built-in rechargeable battery. It was equipped with a USB-mini standard interface for battery recharging and communication with a laptop, PC or tablet.

The development of biosensors within the point-of-care concept assumes that potential users should not possess skills to work with laboratory equipment, such as pipettes. Taking this into consideration, a special sampler adapter for microfluidic chips was developed (Figure 1). This adapter may be used by an unprepared user. The principle of operation of the sampler adapter was as follows. When the chip was placed in the sampler, the chip output channel was located under the clamping ring. The negative pressure in the channel of the chip was created after pressing and releasing the upper part of the sampler. Due to this, the sample was sucked into the microfluidic chip. The sample volume was approximately 45 μl.

Previously, we demonstrated that it was necessary to use an active mixing method for the disposable microfluidic chips, since the passive one did not ensure even distribution of the reagents in the reaction chamber [22]. This uniform distribution was important for the bioluminescent system, since it provided the maximum intensity of luminescence. Therefore, the sampler was used as active mixer. After sampling, periodic clicking on the top of the sampler resulted in active mixing in the chip. It was found that the optimal number of clicks was 10 times. This number provided the most uniform mixing of reagents in reaction chamber, which resulted in higher luminescence intensity level and reproducibility.

3.2. Optimization of microfluidic chip composition and storage conditions

To adapt microfluidic chips for work in the portable luminometer and to improve the storage time of the reagents we optimized concentrations of enzymes and their substrates. It was necessary to achieve luminescence intensity which was high enough to be determined by the SiPM detector.

Four different compositions were used to make microfluidic chips. Microfluidic chips under standard composition (no.1, see Table 1) contained 0.14 μg of luciferase and $52.3 \cdot 10^{-6}$ activity units of NAD(P)H:FMN-oxidoreductase, 0.00044 % of aldehyde, 0.11 mM of NADH. Composition no.2 contained a doubled concentration of aldehyde, composition no.3 in addition to this contained 35% increased concentration of enzymes and composition no.4 in addition to this contained doubled concentration of NADH. Chips of all compositions contained 4 mM of FMN. The chips were tested with the standard laboratory GloMax 20/20 luminometer. The results are shown in Figure 2.

Table 1. Concentrations of enzymes and their substrates in microfluidic chips

	[Aldehyde], %	**Luciferase, μg**	**Oxidoreductase, μU**	**[NADH], mM**
No.1	0.00044	0.14	52.3	0.11
No.2	0.00088	0.14	52.3	0.11
No.3	0.00088	0.19	68.5	0.11
No.4	0.00088	18.90	68.0	0.22

It was demonstrated that the rise of aldehyde concentration led to the increase in the level of luminescence, while the sensitivity to toxin remained the same. The increase in the number of enzymes by 35% led to a sharp increase in the intensity of luminescence by the factor of 5 and a slight loss of sensitivity to the toxic substance. The increase in the concentration of NADH did not affect the intensity of luminescence, but at the same time, increased the sensitivity of the system to the toxic substance. Thus, an excess concentration of aldehyde and NADH in the enzyme system is needed to increase luminescence intensity and sensitivity to toxic compounds. Compositions no. 3 and 4 provided sufficient luminescence intensity to be used with portable luminometer.

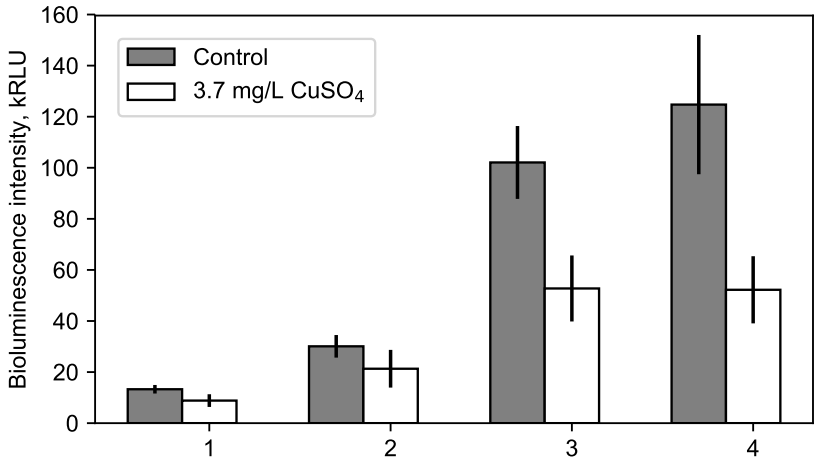

Figure 2. The enzyme system composition in a microfluidic chip was optimized to achieve higher luminescence intensity. Four different compositions were tested. The standard composition (1) contained 0.14 μg of luciferase and $52.3 \cdot 10^{-6}$ activity units of NAD(P)H:FMN-oxidoreductase, 0.00044 % of aldehyde, 0.11 mM of NADH and 4 mM of FMN. Composition (2) contained a doubled concentration of aldehyde, composition (3) in addition contained 35% increased concentration of enzymes and composition (4) in addition contained doubled concentration of NADH.

Changes in the composition of the bioluminescent system did not have a significant effect on the storage time of the chips. With an increased amount of the enzyme, loss of sensitivity to toxin was observed with a storage time of more than one month. A decrease in the luminescence intensity and sensitivity was observed with increasing storage time. Significant reduction of luminescence intensity and sensitivity was observed after 4 months storage at +4 °C. The system completely lost luminescence intensity and sensitivity to toxic substances after 3 months storage at 25 °C. When stored at −18 °C and below the bioluminescent system retained its activity and sensitivity to toxins for 6 months without significant reduction, which was a good result for an immobilized enzyme system [35]. Conventional cell-based biosensors retain their activity only for 1 week [36]. Despite the availability of technologies that allow the bacterial activity to last for several months [37–39], their use requires the availability of certain equipment and specially trained personnel.

3.3. Sensitivity of the portable luminometer

The sensitivity of the SiPM depended on its temperature. To ensure the optimal sensitivity, the system of thermal stabilization was developed. This system was based on a Peltier element, which was controlled by a microcontroller. This system provided thermostabilization of the photodetector at the level of 21 °C. In addition to this, a focusing prism was placed on the detector to improve the light harvest.

To compare the analytical performance of the developed portable SiPM-based luminometer and a bench-scale commercial luminometer GloMax 20/20 we fulfilled a detection of copper (II) sulfate in water by both instruments (Figure 3).

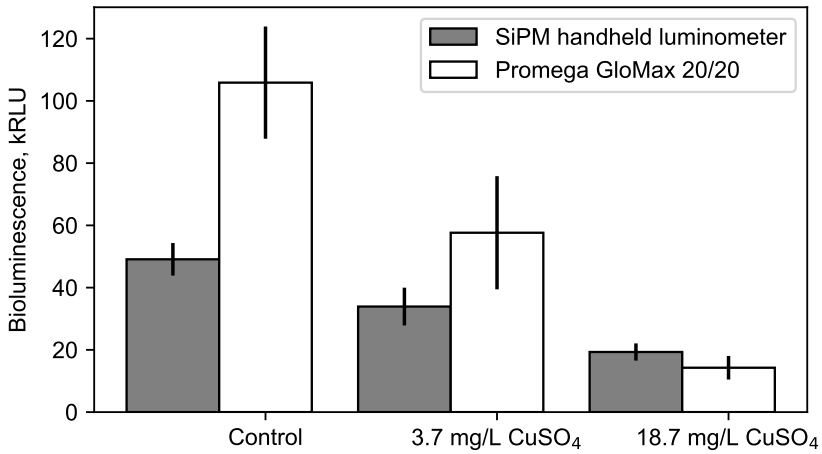

Figure 3. Sensitivity comparison of the biosensor and Promega GloMax 20/20 luminometer by measuring copper(II) sulphate at different concentrations

A control sample (distilled water) and a copper(II) sulphate were measured. After sampling and activation of the bioluminescence reaction, each chip was first placed in GloMax 20/20, where the intensity of the luminescence reached the maximum value. After that, the chip was immediately removed from GloMax 20/20 and placed in a sample compartment of SiPM-base handheld luminometer, where the measurement continued. Such actions were permissible, because upon reaching the maximum intensity of the bioluminescence the signal remained at that level for 10–20 s.

The results of measurements showed that the difference in sensitivity of the developed portable luminometer and GloMax 20/20 had a nonlinear dependence. At low luminescence intensities both luminometers provided comparable level of sensitivity. With increasing signal intensity GloMax 20/20 provided 2 times better resolution than the proposed luminometer. Registered limit of detection for copper(II) sulphate using proposed portable luminometer was 2.5 mg/l.

4. Conclusions

This paper has described the first proofing of concept of handheld biosensor that was designed to detect harmful environmental pollutants in liquid samples. The handheld biosensor consisted of a portable SiPM-based luminometer, disposable microfluidic chips with immobilized bioluminescent enzyme system and a sampler adapter for chips.

Portability and possibility of autonomous operation allow the proposed biosensor to be applied as an early warning system for environmental protection in the field. The compact luminometer can be applied in other areas, for example, medical diagnostics or sanitary, provided there are appropriate microfluidic chips available.

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Conflicts of Interest: The authors of the research K.A.L., I.A.D., V.V.S. are the authors of handheld luminometer studied and described in the article. The E.N.E. is one of the authors of immobilized reagent based on bacterial coupled enzyme system NAD(P)H:FMN-oxidoreductase and luciferase. The funders had no role in the design of

the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

Abbreviations

The following abbreviations are used in this manuscript:

Red+Luc	NAD(P)H:FMN-oxidoreductase and Luciferase;
PMT	Photomultiplier Tube;
SiPM	Silicon Photomultiplier;
FMN	Flavin Mononucleotide;
NADH	Nicotinamide Adenine Dinucleotide;
CAD	Computer-aided Design.

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